

A GUIDE TO SELECTING

Control and Blocking Reagents



OPTIMIZE YOUR EXPERIMENTAL PROTOCOLS

Optimize your experimental protocols with control, diluent and blocking reagents.

When developing an immunotechnique, it is important to consider result analysis. The addition of the correct blocking reagents and experimental controls can improve assay performance and interpretation. The inclusion of positive and negative controls is important when generating data for publication.

Use this guide to learn more about the selection of appropriate control, diluent and blocking reagents in a range of immunotechniques to:

- Reduce background.
- Troubleshooting off-target signals.
- Facilitate analysis.

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Flow cytometry

Problem	Solution	Indicated product
Background from antibodies binding Fc receptors	Block Fc receptors.	Normal serum from the host of the labeled antibody
	Use F(ab') ₂ format secondary antibody to avoid entrapment by Fc receptors.	F(ab') ₂ secondary antibodies
Confirmation that primary antibody binding is due to antigen specificity	Use an isotype negative control (non-specific IgG from the same species as the primary antibody) to demonstrate specific binding of the primary antibody.	ChromPure™ purified proteins
Confirmation that secondary antibody does not contribute to off-target signal	Use an isotype negative control (conjugated non-specific IgG from the same species as the secondary antibody) to demonstrate specific binding of the secondary antibody.	ChromPure™ purified proteins
Short on time and no directly conjugated primary available	Fab label your primary antibodies prior to incubation with sample.	FabuLight™



Western blotting

Problem	Solution	Indicated product
Background (non-specific signal obscuring bands of interest)	Use appropriate blocking reagent to block membrane prior to incubating with primary antibody.	Normal serum (5% v/v) from the host species of the labeled antibody, or BSA (5% w/v)(IgG-free and protease- free)
	Avoid using milk or BSA if using primary antibodies derived from goat, horse or sheep. Bovine IgG may interact with the antibody due to homologous epitopes of closely related species.	Normal serum (5% v/v) from the host species of the labeled antibody
Interference from reduced immunoprecipitating (IP) antibody when detecting 50 or 25 kDa proteins	To avoid detecting IP antibody heavy chains at 50 kDa, probe blot with conjugated anti-Light Chain specific antibody.	Anti-Light Chain antibodies See online or catalog for more information.
	To avoid detecting IP antibody light chains at 25 kDa, probe blot with conjugated anti-IgG. Fc fragment <i>after blocking with monovalent Fab fragment anti-Fc.</i>	Anti-Fc antibody Fab fragments

To confirm reporter enzyme activity, add a small sample of conjugated secondary directly to substrate and observe expected reaction.



ELISA

Problem	Solution	Indicated product
Background	Use an appropriate volume of blocking reagent to completely block wells prior to incubating with the primary antibody.	Normal serum (5% v/v) from the host species of the labeled antibody, or BSA (5% w/v)(IgG-free and protease-free)
No signal	Use a positive control to demonstrate activity of the labeled secondary antibody, coat with primary antibody isotype and detect directly with secondary antibody	ChromPure™ purified proteins

Cautions for blocking with bovine serum albumin (BSA) or milk:

Bovine serum albumin (BSA) and dry milk sometimes contain bovine IgG. **With the exception of Jackson ImmunoResearch Bovine Anti-Goat IgG**, many secondary antibodies such as Anti-Bovine, Anti-Goat, and Anti-Sheep will react with bovine IgG. Therefore, use of BSA or dry milk for blocking or diluting may significantly increase background and/or reduce antibody titer. For blocking, use normal serum (5% v/v) from the host species of the labeled secondary antibody.

For additional information on IgG- and Protease-free BSA from Jackson ImmunoResearch, see page 10.

Immunohistochemistry, Immunocytochemistry, Immunofluorescence

Problem	Solution	Indicated Product
Confirmation that primary antibody binding is due to antigen specificity	Use an isotype negative control (non-specific IgG from the same species as the primary antibody) to demonstrate specific binding of the primary antibody.	ChromPure™ proteins
Background (general)	Block endogenous binding sites which may interact with experimental reagents.	Normal serum from the host of the labeled antibody.
	Dilute antibodies in buffer without carrier proteins. Centrifuge working dilution to remove aggregates.	Correct diluent and preparation, e.g. PBS/ Tween 20
Background (homologous Ig recognition)	Use a cross-adsorbed secondary antibody with minimal cross-reactivity to the cell or tissue species being analyzed.	Min X secondary antibodies
	Block endogenous immunoglobulins.	Fab fragments
Multiple labeling of primary antibodies from same host species (e.g. mouse on mouse)	Utilize Fab fragments in suggested protocols to accomplish multiple labeling (see online).	Fab fragments
	Immunolabel primary antibody prior to incubation.	FabLight™
Endogenous enzymes	Inactivate endogenous peroxidases with hydrogen peroxide.	Hydrogen peroxide
	Inactivate endogenous phosphatases with levamisole.	Levamisole
Endogenous biotin	Block endogenous biotin.	Incubate with streptavidin, followed by free biotin.
Ionic or hydrophobic interactions	Include detergent in buffers, optimize salt concentrations and pH.	Tween 20 and/or Triton X-100

Immunoreagents

Jackson ImmunoResearch offers a wide variety of immunoreagents designed to improve assay performance and ease of analysis. The immunoreagents below are highlighted within this guide. Please visit our website or reference our catalog for more information on the extensive range of secondary antibodies and conjugates manufactured at Jackson ImmunoResearch.

- **Normal Serums**
- **ChromPure™ Purified Proteins from Normal Serums**
- **Monovalent Fab Fragment Affinity-Purified Antibodies**
- **IgG-Free, Protease-Free Bovine Serum Albumin**

If you require additional support with antibody selection or are experiencing problems with your experiment, please contact our technical team at tech@jacksonimmuno.com.



Normal Serums

Normal serums are obtained from non-immunized animals, and consequently do not detect any specific antigen. Normal serum diluted to 5% (v/v) in PBS (or comparable buffer) is strongly recommended as a blocking agent to reduce background from non-specific, conserved-sequence, and/or Fc-receptor binding. Best results are obtained with diluted normal serum from the same host as the labeled antibody, used as a separate incubation step before addition of the primary antibody.

ChromPure™ Purified Proteins from Normal Serums

ChromPure™ proteins are primarily used as experimental controls for either primary or secondary antibodies. They may also be used as blocking reagents for Western blotting, IHC and IF.

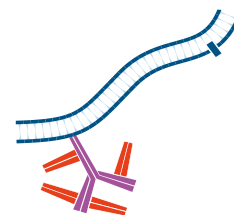
ChromPure™ proteins are derived from the serum of non-immunized animals and do not recognize any known antigens. They are prepared using a variety of chromatographic techniques to yield material with no contaminating molecules observed up to a concentration of 20 mg/ml, making them ideal for use as experimental controls for the most sensitive of assays.

ChromPure™ proteins are available in a variety of formats for many species, including whole immunoglobulin, F(ab')₂ and Fab fragments. Human IgM, serum IgA and other proteins are also available. Jackson ImmunoResearch carries an extensive range of conjugates for this product line, including a range of fluorescent dyes and reporter enzymes, allowing the isolation of signal derived from non-specific interactions.

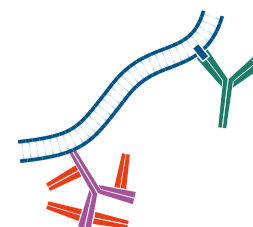
Monovalent Fab Fragment Affinity-Purified Antibodies

Fab fragments can be used to block endogenous immunoglobulins to reduce background staining and to double label primary antibodies from the same host species. The following example shows how Fab fragments can be used to block endogenous immunoglobulins when using mouse primary antibodies on mouse tissue.

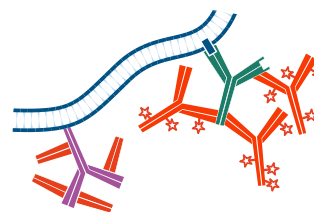
Example: Use unconjugated Fab fragments to block endogenous immunoglobulins to avoid off target signal.



1. Samples which express endogenous IgG may cause off target staining. After blocking with normal serum, incubate with an excess of unconjugated Fab antibody, in this case Fab fragment Goat Anti-Mouse IgG (H+L). Wash.



2. Incubate with primary antibody (Mouse Anti-Antigen X). Wash.

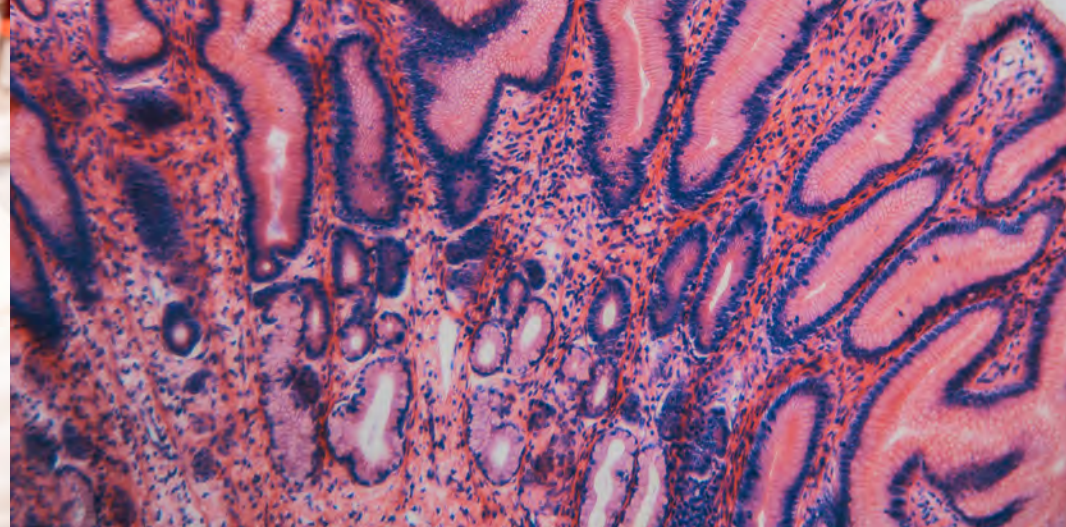


3. Incubate with conjugated secondary antibody, in this case Alexa Fluor® 647-Goat Anti-Mouse IgG (H+L). Wash.

Key

	Endogenous Immunoglobulin		Antigen X
	Mouse Anti-Antigen X		Alexa Fluor® 647
	Goat Anti-Mouse IgG (H+L)		Fab fragment Goat Anti-Mouse IgG (H+L).





IgG-Free, Protease-Free Bovine Serum Albumin

Bovine serum albumin (BSA) is used extensively as a carrier protein to dilute antibodies and as a general protein blocking agent in immunoassays and immunodetection protocols. If BSA is the desired diluent or blocking reagent for your assay it's important to use BSA that is suitable for the purpose.

Most BSA products, including those marketed as having no detectable IgG, are contaminated with low levels of bovine IgG.

Bovine IgG shares many epitopes with goat, sheep and horse IgG, and can become a target for secondary antibodies directed against those species (an exception is bovine anti-goat IgG). This may occur with other antibodies that cross-react with bovine IgG as well. The interaction may result in loss of antibody activity, and/or increased background. The background may derive from sticky, soluble immune complexes or from non-specific binding from contaminating bovine IgG attracting cross-reacting labeled secondary antibodies.

Jackson ImmunoResearch Bovine Serum Albumin is IgG-free and protease-free. It does not contain contaminating IgG, which alleviates common immunoassay problems associated with many commercial high purity preparations of BSA. IgG-free BSA is supplied as a pure protein, freeze-dried from deionized water in 10 g, 50 g, and 250 g pack sizes.

References

- Alberts B et al (1994) Molecular biology of the Cell. 3rd Ed. Garland Press. London.
- Kalyuzhny A (2016) Immunohistochemistry - Essential Elements and Beyond. Springer International Publishing Switzerland.
- Mann, M. & Jensen, O.N. (2003) Proteomic analysis of post-translational modifications. Nat. Biotechnol. 21, 255-261.



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